



Intense exercise potentiates oxidative stress in striatum of reserpine-treated animals

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ARTICLE INFO

Article history:

Received 16 May 2008

Received in revised form 7 November 2008

Accepted 25 November 2008

Available online 10 December 2008

Keywords:

Intense exercise

Orofacial dyskinesia

Reserpine

Oxidative stress

Neurotoxicity

ABSTRACT

Regular physical activity exerts beneficial effects for mental and physical health, but an intense exercise can cause oxidative stress (OS) in dopaminergic regions and intensify the harmful effects of reserpine. Reserpine-induced neurotoxicity can be accessed by behavioral and biochemical evaluations. The objective of this study was to examine the effect of a gradual intensifying exercise program on an animal model of oxidative stress. Male rats were submitted to swimming sessions (1 h/day, for eleven weeks), and they were loaded gradually during the adaptation period (two weeks) with a weight corresponding to 1–7% of their body weight tied to their back. After the last training, the animals were treated with two doses of vehicle or reserpine (1 mg/kg-sc), an agent that induces orofacial dyskinesia. After behavioral evaluations, the striatum was dissected for enzymatic and biochemical assays. Development of cardiac hypertrophy demonstrated the effectiveness of the physical training. The gradual intense exercise and reserpine increased lipid peroxidation and striatal catalase activity. The results confirm the importance of catalase activity in orofacial dyskinesia which can be related to lipid peroxidation in striatal dopaminergic brain tissue. These results indicate that intense exercise can have some deleterious effect on striatal dopaminergic system.

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1. Introduction

Regular physical exercise exerts many benefits for mental and physical health, reducing disease incidence, improving cognitive processes and increasing resistance to brain injury (Griesbach et al., 2004; Ogonovszky et al., 2005; Radak et al., 2001). Accumulated evidence has shown that regular physical activity may increase antioxidant defenses against brain oxidative damage (Radak et al., 2006; Somani et al., 1995) and induce resistance to oxidative stress (OS) (Banerjee et al., 2003; Ji, 1999) promoting antiapoptotic effects (Toldy et al., 2005). In this sense, moderate physical activity can offer protection against seizures (Setkovicz and Mazur, 2006; Dubow and Kelly, 2003), neurodegenerative diseases (Howells et al., 2005; Kiraly and Kiraly, 2005; Smith and Zigmund, 2003; Sutoo and Akiyama, 2003; Sasco et al., 1992), and alleviate symptoms of anxiety or depression (Martinsen et al., 1985).

Literatures data about the effect of regular exercise on brain oxidative stress (OS) and brain functioning have been conflicting (Radak et al., 2001; Suzuki et al., 1983). Indeed, part of literature supports a beneficial influence while other portion supports a deleterious effect of exercise. Considering different forms and intensity of physical activity, some research groups have demonstrated a relationship between exhaustive exercise and the increased generation of reactive oxygen species (ROS)

in different biological tissues (Bejma et al., 2000; Reid et al., 1992; Sastre et al., 1992). In addition, excessive physical activity increased abruptly may generate psychological symptoms that resemble clinical depression (Armstrong and Van Heest, 2002; Budgett, 1998; Petibois et al., 2003) and a stress-related condition that may alter physiological and immunological functions associated with biochemical abnormalities (Angeli et al., 2004).

Of particular importance, the brain is more susceptible to OS when compared to other organs or systems, because it contains high content of unsaturated membrane lipids, excitotoxic amino acids, low levels of antioxidant defenses and autoxidizable neurotransmitters (Halliwell and Gutteridge, 1999). Free radicals are formed in the central nervous system (CNS) as part of normal metabolic processes (Halliwell, 1992), but when the production of ROS exceeds the ability of the antioxidant system to eliminate them, oxidative damage occurs (Jenkins and Goldfarb, 1993). This subject is extremely important since the brain is exposed throughout life to OS and there is evidence of the involvement of free radicals, over production in the pathophysiology of several diseases of the nervous system (Gilgun-Sherki et al., 2001). Among these diseases are included Alzheimer's (Pappolla et al., 1998), Parkinson's (Offen et al., 1999; Ziv et al., 1994), Huntington's (Browne et al., 1997), schizophrenia (Mahalik and Scheffer, 1996) and tardive dyskinesia (Tsai et al., 1998).

Reserpine is a monoamine depletor which prevents the storage of dopamine (DA) in neuronal synaptic vesicles. Reserpine interferes with the vesicular monoamine transporter (VMAT), causing an increase in

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cytosolic DA that can be oxidatively metabolized by MAO. This accelerated DA metabolism can lead to the formation of reactive metabolites and hydrogen peroxide, which can be associated with the OS process in dopaminergic neurons (Abílio et al., 2003a; Bilska and Dubiel, 2007; Burger et al., 2003; Naidu et al., 2004). Furthermore, autoxidation of DA produce *O*-quinone aminochrome that can suffer one-electron reduction to form the leukoaminochrome *O*-semiquinone radical, which is thought to be one of the major sources of endogenous reactive species involved in the degenerative processes (Fuentes et al., 2007; Paris et al., 2001, 2005; Segura-Aguilar et al., 2002). Particularly, the brain basal ganglia are rich in monoamines and therefore more vulnerable to free radical damage leading to OS (Lohr et al., 2003).

Experimentally, reserpine has been used by various laboratories to study movement disorder related to OS and neurodegenerative disease (Abílio et al., 2003b; Bilska and Dubiel, 2007; Castro et al., 2006; Dutra et al., 2002; Naidu et al., 2004; Peixoto et al., 2005). Recently, our laboratory has shown the beneficial effects of moderate chronic exercise on behavioral and biochemical parameters in an animal model of reserpine-induced OS (Teixeira et al., 2008). In view of the conflicting data found in the literature, the aim of the present study was to evaluate behaviorally and biochemically the influence of a gradual intense physical activity on brain susceptibility to OS, either alone or in association with reserpine, an OS-inductor. Particular attention was given to catalase in view of the recent studies of Abílio et al. (2004), which have shown an association between OD and catalase activity.

2. Method

2.1. Drugs

Reserpine (methyl reserpate 3,4,5-trimethoxybenzoic acid ester-Sigma Chemical) was dissolved in glacial acetic acid and then diluted to a final concentration of 0.5% acetic acid with distilled water. The vehicle consisted of a 0.1% acetic acid solution. These solutions were injected subcutaneously (sc), at a volume of 1.0 mL/kg body weight.

2.2. Animals

The experiment was conducted with 32 male Wistar rats weighing between 160 and 180 g at the start of the experiments. Groups of eight animals were kept in Plexiglas cages with free access to food and water in a room with controlled temperature ($23\text{ }^{\circ}\text{C}\pm 1\text{ }^{\circ}\text{C}$) and on a 12 h-light/dark cycle with lights on at 7:00 a.m. The number of animals used was the minimum to obtain relevant results and they were maintained and used in accordance to the guidelines of the Brazilian Association for Laboratory Animal Science (COBEA), following international norms of care and animal maintenance. The rats were randomly assigned to four groups: sedentary-control (SC), sedentary-reserpine (SR), exercise-control (EC), exercise-reserpine (ER).

2.3. Training protocol and experimental procedure

All exercised rats were subjected to swimming exercise in a plastic container (depth 45 cm) under continuous supervision, with the water temperature set to $34\text{ }^{\circ}\text{C}\pm 1\text{ }^{\circ}\text{C}$, 1 h/day, 5 times per week during 11 weeks. The duration of swimming was initially 15 min and was gradually increased by 15 min/day until reach a maximal of 60 min that was maintained during all the training period. The masses of the weights, which were fixed at the back of the rats, were increased gradually during two weeks, until a maximal of 7% of rats body weight (Chaumont et al., 2001).

One day after the last training session, all the animals received an injection of vehicle (0.1% acetic solution) or reserpine solution (1 mg/kg body weight in 0.1% of acid acetic solution), subcutaneously, for 3 days every other day, totaling two injections. The animals' weight was recorded weekly during the experiment.

2.4. Training verification

Immediately after the behavioral evaluations (see Sections 2.5 and 2.6) all the rats were sacrificed by decapitation and the heart was removed and weighed. The heart weight (mg)/body weight (g) (HW/BW) ratio was used as an index of cardiac hypertrophy (Tharp and Carson, 1975).

2.5. Behavioral testing

On the fourth day (between 22 and 24 h after the last reserpine or vehicle injection), all the rats were observed for the quantification of orofacial dyskinesia (OD): The animals were placed individually in cages ($20\times 20\times 19\text{ cm}$) containing mirrors under the floor to allow behavioral quantification when the animal was faced away from the observer. To quantify the occurrence of oral dyskinesia, the incidence of vacuous chewing movements (VCM) frequency and the duration of facial twitching (FT) was recorded during 5 min after a period of 2 min adaptation (hand operated counters were employed). Observers were blind to the drug treatments.

The behavioral experiments were conducted between 09:00 and 11:00 a.m.

2.6. Tissue preparation and oxidative stress parameters

Immediately after the behavioral evaluation (between 26 and 28 h after the last reserpine or vehicle injection), the rats were decapitated; the brains were removed and put on ice, cut coronally at the caudal border of the olfactory tubercle. The striatum was dissected from the anterior part, and separated into two parts. The right striatum was homogenized in 10 volumes (w/v) of 0.1 M Tris-HCl, pH 7.4, centrifuged at $3000\times g$ for 10 min and the supernatants were used for lipid peroxidation and catalase activity. Lipid peroxidation was determined by measuring accumulation of thiobarbituric acid reactive substances (TBARS) as described by Ohkawa et al. (1979). Catalase was quantified by measuring the disappearance of H_2O_2 at 240 nm (Aebi et al., 1995).

2.7. Statistical analysis

All the data were analyzed by a two-way ANOVA (2 (sedentary/exercise) \times 2 (control/reserpine)) followed by Duncan's multiple range test, when appropriated (Siegel, 1956).

3. Results

Verification of training program: Heart weight (mg), final body weight (g), heart weight/body weight ratios (heart hypertrophy index) are shown in Table 1. Two way ANOVA revealed a significant main effect of intense exercise on heart weight [$F(1,20)=56.90$, $P<0.001$] and heart weight/body weight ratios [$F(1,20)=57.21$, $P<0.001$]. Univariate ANOVA followed by Duncan's multiple range test revealed that exercised groups (EC and ER) showed increase of

Table 1

Mean values of heart weight, final body weight and heart weight/body weight ratios for sedentary (S) and exercised (E) rats, treated with vehicle or reserpine (C and R, respectively)

Group	Heart weight (mg)	Final body weight (g)	Ratio (mg/g)
SC	972.80 \pm 43.0	367.8 \pm 5.8	2.64 \pm 0.10
SR	1027.63 \pm 10.6	388.0 \pm 4.5	2.65 \pm 0.10
EC	1281.10 \pm 26.7*	378.5 \pm 6.8	3.38 \pm 0.10*
ER	1247.30 \pm 47.1*#	374.5 \pm 7.8	3.33 \pm 0.09*#

Data (mean \pm S.E.M.), ($n=8$) were analyzed by two-way ANOVA followed by Duncan's test. * ($P<0.001$) difference from sedentary-control group (SC); # ($P<0.001$) difference from sedentary-reserpine group (SR).

heart weight and of heart hypertrophy (organ-to-body weight ratio), when compared with sedentary groups (SC and SR), indicating the effectiveness of the exercise protocol. Body weight was not modified by physical training or reserpine administration.

Orofacial movements: The effects of reserpine treatment on orofacial movements in rats are shown in Fig. 1. Two way ANOVA revealed a significant main effect of reserpine [$F(1,28)=178.31$; $P<0.001$] and a significant reserpine \times exercise interaction [$F(2,28)=6.92$; $P<0.05$] for VCM frequency (Fig. 1A). Univariate ANOVA followed by Duncan's multiple range test revealed that sedentary and exercised rats treated with reserpine (SR and ER) displayed an increase in VCM when compared to control groups (SC and EC). In fact, ER group showed VCM frequency significantly higher than that of both reserpine-treated sedentary (SR) and vehicle-treated (EC) exercised rats.

For duration of FT, two way ANOVA yielded a significant main effect of reserpine [$F(1,28)=272.68$; $P<0.001$] (Fig. 1B). Univariate analysis, followed by Duncan's multiple range test, revealed that sedentary and exercised rats treated with reserpine (SR and ER) displayed an increase in FT duration when compared to control group (SC). In fact, the duration of FT of reserpine-treated exercised rats (ER) was significantly higher than that of exercised rats (EC), and similar to reserpine-treated sedentary group (SR).

3.1. Biochemical analysis

The results of intense exercise and reserpine treatment on TBARS levels and catalase activity in brain striatum are shown in Fig. 2. Two way ANOVA of TBARS levels revealed a significant main effect of exercise [$F(1,28)=132.54$; $P<0.001$], reserpine treatment [$F(1,28)=142.47$; $P<0.001$] and exercise \times reserpine interaction [$F(2,28)=37.10$; $P<0.001$] (Fig. 2A). Post hoc comparisons by Duncan's multiple range test indicated a significant increase on TBARS levels in ER group when compared to the other groups. The reserpine administration (SR) and the exercise (EC) caused *per se* a significant increase in TBARS levels, when compared to control group (SC).

Two way ANOVA revealed a significant main effect of exercise and reserpine on striatal catalase activity, [$F(1,28)=4.32$, $P<0.046$] and [$F(1,28)=39.68$, $P<0.001$] respectively, (Fig. 2B). Univariate ANOVA followed by Duncan's multiple range test revealed that sedentary and exercised rats treated with reserpine showed a significant increase of catalase activity when compared to the control group (SC). In fact, ER group showed catalase activity significantly higher than

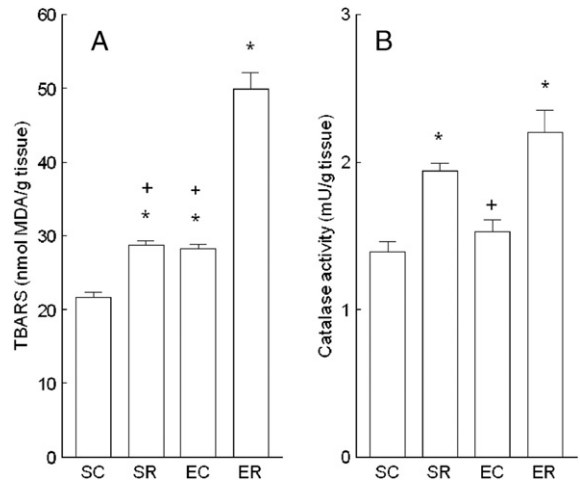


Fig. 2. Effects of reserpine administration (1.0 mg/kg sc every other day, for 3 days) (SR) or vehicle (SC) on sedentary rats ($n=16$) and rats submitted to intense exercise ($n=16$) (ER) or (EC) on TBARS levels (A) and catalase activity (B) in the striatum. Data (mean \pm S.E.M.) were analyzed by two-way analysis of variance followed by Duncan's test. *Indicates a significant difference from SC group for $P<0.05$, * indicates a significant difference from ER group for $P<0.001$.

that of reserpine-treated sedentary (SR) and vehicle-treated (EC) exercised rats.

4. Discussion

Recently we have showed the beneficial effects of chronic physical exercise (moderate intensity) in an OS animal model, suggesting beneficial effects in neurological diseases associated with orofacial movements (Teixeira et al., 2008). Here we have studied a gradual increasing in the exercise intensity because variations in the frequency and content of the physical exercise can have an influence on brain functions, leading to various disturbances, which can be measured by behavioral, biochemical and genetic modifications (Kleim et al., 2003; Sarbadhikari, 1995). Interestingly and similar to other researchers, we did not observe variation in the body weight of the exercised animals compared to the controls at the end of the experiment (Gündüz et al., 2004; Radak et al., 2001). The reasons for the absence of an effect of exercise on final body weight remains unknown, but it can be related to change in the proportion of muscular and fat tissue in the sedentary and trained groups.

The primary factor involved in reserpine-induced vacuous chewing behaviour is related to dopamine depletion and reduced activation of D_2 receptors (Neisewander et al., 1991, 1994). In line with this, literature data have indicated that dopamine plays an important role in motor activation (Freed and Yamamoto, 1985) and an increase in dopamine metabolism was observed in several brain regions of rats during physical activity (Meeusen and De Meirleir, 1995). Of particular interest for the animal model chosen here, literature data have indicated the participation of an elevation in dopamine metabolism with orofacial movement disorders in reserpine-induced animal models and in human diseases (Abílio et al., 2003a; Burger et al., 2003; Cadet and Kahler, 1994; Faria et al., 2005; Naidu et al., 2003; Raghavendra et al., 2001; Sagara, 1998; Singh et al., 2003; Sussman et al., 1997). In this sense, the reserpine is an OS inductor, mainly by preventing the storage of dopamine (DA) in neuronal vesicles via inhibition of the vesicular monoamine transporter (VMAT). This causes an increase in the catabolism of cytosolic DA (by Monoamine oxidase), which can contribute to accelerate the mechanism(s) leading to OS (Abílio et al., 2003a; Bilzka et al., 2007; Burger et al., 2003; Naidu et al., 2004). In line with this, this monoamine depletor reduces the uptake of DA, increasing its availability in the cytosol of dopaminergic neurons. This cytosolic DA can be oxidized to *O*-quinone aminochrome

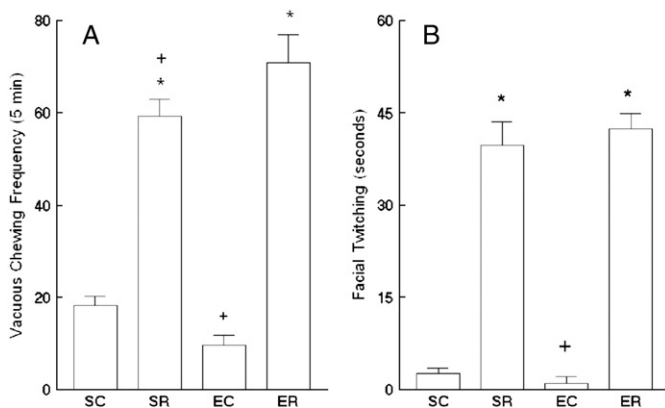


Fig. 1. Effects of reserpine administration (1.0 mg/kg sc every other day, for 3 days) (SR) or vehicle (SC) on sedentary rats ($n=16$) and rats submitted to intense exercise ($n=16$) (ER) or (EC) on vacuous chewing frequency (A) and facial twitching duration (B). Data (mean \pm S.E.M.) were analyzed by two-way analysis of variance followed by Duncan's test. *Indicates a significant difference from SC group for $P<0.001$, * indicates a significant difference from ER group for $P<0.001$.

stimulating one of the major sources of reactive species involved in the degenerative processes (Paris et al., 2001, 2005; Segura-Aguilar et al., 2002).

In our previous study, we have demonstrated that moderate chronic exercise displayed a beneficial effect by partially preventing the increase in FT induced by reserpine (Teixeira et al., 2008). Here, in contrast, we observed that the reserpine treatment increased VCM and FT and that gradual intense physical activity did not alter these parameters. Exercise and reserpine treatments increased TBARS when compared to control group in striatal tissue. In fact, the gradual increase in the intensity of exercise was not enough to prevent the development of lipid peroxidation in the striatal region, which was more evident when in association with reserpine (OS inductor). Together, these results shown that the effects of intense exercise and reserpine treatment may result in an increase of dopamine metabolism. In fact, the pro-oxidant effect of the association of reserpine with exercise was observed by the increase of TBARS levels and catalase activity. The development of OD may be related to this striatal oxidative damage, which is in line with the view of literature data showing that catalase activity exerts a critical role in the development of OD and OS (Abílio et al., 2004; Faria et al., 2005). Similarly, Margonis et al. (2007) have explained an increase of catalase activity by a compensatory mechanism involved in the scavenging of free radicals after exercise.

Recently, Acikgoz et al. (2006) showed that acute exhaustive exercise did not change antioxidant defenses in frontal cortex and striatum, suggesting that exercise does not induce significant OS, and their results are not contradictory to those presented here because our experimental paradigm was performed chronically. In line with this, different researchers have submitted rats to moderate chronic exercise and observed an increase of catalase activity in brain and other tissues, relating this effect to an increase of antioxidant defenses (Devi and Kiran, 2004; Gündüz et al., 2004; Liu et al., 2000; Somani et al., 1995). Here we have found that the intense exercise protocol associated to reserpine treatment caused an increase in striatal catalase activity. In this sense, the increase in catalase activity can be a compensatory mechanism involved in the detoxification of reactive species in dopaminergic brain regions that can be related to a higher consumption of oxygen after chronic intense exercise (Jenkins, 1988; Sjödin et al., 1990) and to a greater generation of dopamine-quinones by reserpine administration (Fuentes et al., 2007). Various researchers have determined the effects of exercise on the antioxidant defenses in different tissues, including in brain regions (Somani et al., 1995; Ogonovszky et al., 2005; Acikgoz et al., 2006). The results presented here demonstrate for the first time that chronic and gradual intense exercise can change OS and antioxidant defenses in striatum which can be involved in movement disorders. These findings may be particularly important in clinical situations, when the OS and the neurotoxicity are closely involved.

In conclusion, the results presented here indicated that, in general, gradual chronic intense exercise had pro-oxidant effect by itself (as assessed by TBARS in striatum). In addition, when it was associated with reserpine treatment, intense physical exercise caused further increase in striatal TBARS and orofacial dyskinesia (OD), suggesting that OS is an important factor in the development of OD.

Acknowledgments

The financial support from the CAPES and the CNPq is gratefully acknowledged.

References

- Abílio VC, Vera JAR, Ferreira LSM, Duarte CRM, Martins CR, Torres-Leite D, et al. Effects of melatonin on behavioral dopaminergic supersensitivity. *Life Sci* 2003a;72:3003–15.
- Abílio VC, Araujo CC, Bergamo M, Calvente PR, D'Almeida V, Ribeiro R, Frussa-Filho R. Vitamin E attenuates reserpine-induced oral dyskinesia and striatal oxidized glutathione/reduced glutathione ratio (GSSG/GSH) enhancement in rats. *Prog Neuro-Psychopharmacol Biol Psychiat* 2003b;27:109–14.
- Abílio VC, Silva RH, Carvalho RC, Grassl C, Calzavara MB, Registro S, et al. Important role of striatal catalase in aging and reserpine-induced oral dyskinesia. *Neuropharmacology* 2004;47:263–72.
- Acikgoz O, Aksu I, Topcu A, Kayatekin BM. Acute exhaustive exercise does not alter lipid peroxidation levels and antioxidant enzyme activities in rat hippocampus, prefrontal cortex and striatum. *Neurosci Lett* 2006;406:148–51.
- Aebi U, Chiu W, Milligan R. Role of catalase on antioxidative defenses. *J Struct Biol* 1995;2:117–8.
- Armstrong L, Van Heest J. The unknown mechanism of the overtraining syndrome: clues from depression and psychoneuroimmunology. *Sports Med* 2002;32:185–209.
- Angeli A, Minetto M, Dovic A, Paccotti P. The overtraining syndrome in athletes: a stress-related disorder. *J Endocrinol Invest* 2004;27:603–12.
- Banerjee AK, Mandal A, Chanda D, Chakraborti S. Oxidant, antioxidant and physical exercise. *Mol Cell Biochem* 2003;253:307–12.
- Bejma J, Ramirez P, Jilka. Free radical generation and oxidative stress with aging and exercise: differential effects in the myocardium and liver. *Acta Physiol Scand* 2000;169:343–51.
- Bilska A, Dubiel M. Alpha-lipoic acid differently affects the reserpine-induced oxidative stress in the striatum and prefrontal cortex of rat brain. *Neuroscience* 2007;146:1758–71.
- Browne SE, Bowling AC, MacGarvey U, Baik MJ, Berger SC, Muqit MM, et al. Oxidative damage and metabolic dysfunction in Huntington's disease: selective vulnerability of the basal ganglia. *Ann Neurol* 1997;41:646–53.
- Budgett R. Fatigue and underperformance in athletes: the overtraining syndrome. *Br J Sports Med* 1998;32:107–10.
- Burger M, Alves A, Callegari L, Athayde FR, Nogueira CW, Zeni G, Rocha JBT. Ebselen attenuates reserpine-induced orofacial dyskinesia and oxidative stress in rat striatum. *Prog Neuro-Psychopharmacol Biol Psychiat* 2003;27:135–40.
- Cadet JL, Kahler LA. Free radical mechanisms in schizophrenia and tardive dyskinesia. *Neurosci Biobehav* 1994;18:457–67.
- Castro JPMV, Frussa-Filho R, Fukushiro DF, Silva RH, Medrano WA, Ribeiro R, Abílio VC. Effects of baclofen on reserpine-induced vacuous chewing movements in mice. *Brain Res Bull* 2006;68:436–41.
- Chaumont SB, Maupoil V, Lahet JJ, Berthelot A. Effect of exercise training on metallothionein levels of hypertensive rats. *Med Sci Sports Exer* 2001;33:724–8.
- Devi SA, Kiran TR. Regional responses in antioxidant system to exercise training and dietary vitamin E in aging rat brain. *Neurobiol Aging* 2004;25:501–8.
- Dubow JS, Kelly JP. Epilepsy in sports and recreation. *Sports Med* 2003;33:499–516.
- Dutra RC, Andrezza AP, Andreatini R, Tufik S, Vital MABF. Behavioral effects of MK-801 on reserpine-treated mice. *Prog Neuro-Psychopharmacol Biol Psychiat* 2002;26:487–95.
- Faria RR, Abílio VC, Grassl C, Chinen CC, Negrão LTR, Castro JPMV, et al. Beneficial effects of vitamin C and vitamin E on reserpine-induced oral dyskinesia in rats: critical role of striatal catalase activity. *Neuropharmacology* 2005;48:993–1001.
- Freed C, Yamamoto B. Regional brain dopamine metabolism: a marker for the speed, direction and posture of moving animals. *Science* 1985;229:62–5.
- Fuentes P, Paris I, Nassif M, Caviedes P, Segura-Aguilar S. Inhibition of VMAT-2 and DT-diaphorase induced cell death in a substantia nigra-derived cell line: an experimental cell model for dopamine toxicity studies. *Chem Res Toxicol* 2007;20:776–83.
- Gilgun-Sherki Y, Melamed E, Offen D. Oxidative stress induced-neurodegenerative diseases: the need for antioxidants that penetrate the blood barrier. *Neuropharmacology* 2001;40:959–75.
- Griesbach GS, Hovda DA, Molteni R, Wu A, Gomes-Pinilla F. Voluntary exercise following traumatic brain injury: brain-derived neurotrophic factor upregulation and recovery of function. *Neuroscience* 2004;125:129–39.
- Gündüz F, Sentürk UK, Kuru O, Aktekin B, Aktekin MR. The effect of one year's swimming exercise on oxidant stress and antioxidant capacity in aged rats. *Physiol Res* 2004;53:171–6.
- Halliwell B. Reactive oxygen species and the central nervous system. *J Neurochem* 1992;59:1609–23.
- Halliwell B, Gutteridge JMC. Free radicals in biology and medicine. Third ed. New York: Oxford University Press; 1999. p. 645–60.
- Howells FM, Russell VA, Mabandla MV, Kellaway LA. Stress reduces neuroprotective effect of exercise in a rat model of Parkinson's disease. *Behav Brain Res* 2005;165:210–20.
- Jenkins R. Free Radical chemistry: relationship to exercise. *Sports Med* 1988;5:156–70.
- Jenkins R, Goldfarb A. Introduction: oxidant stress, aging, and exercise. *Med Sci Sports Exer* 1993;25:210–2.
- Ji LL. Antioxidants and oxidative stress in exercise. *Proc Soc Exp Biol Med* 1999;222:283–92.
- Kiraly MA, Kiraly SJ. The effect of exercise on hippocampal integrity: review of recent research. *Int J Psychiatry Med* 2005;35:75–89.
- Kleim JA, Jones TA, Schallert T. Motor enrichment and the induction of plasticity before or after brain injury. *Neurochem Res* 2003;28:1757–69.
- Liu J, Yeo HC, Övervik-Douki E, Hagen T, Doniger SJ, Chu DW, et al. Chronically and acutely exercised rats: biomarkers of oxidative stress and endogenous antioxidants. *J Appl Physiol* 2000;89:21–8.
- Lohr JB, Kuczenski R, Niculescu AB. Oxidative mechanisms and tardive dyskinesia. *CNS Drugs* 2003;17:47–62.
- Mahadik SP, Scheffer RE. Oxidative injury and potential use of antioxidants in schizophrenia. *Prostagl Leuk Ess Fatty Acids* 1996;55:45–54.
- Margonis K, Fatouros IG, Jamurtas AZ, Nikolaidis MG, Douroudos I, Chatziniolaou A, et al. Oxidative stress biomarkers responses to physical overtraining: implications for diagnosis. *Free Radic Biol Med* 2007;43:901–10.
- Martinsen EW, Medhus A, Sandvik L. Effects of aerobic exercise on depression, a controlled study. *Br Med J (Clin res ed)* 1985;291:109.
- Meeusen R, De Meirleir K. Exercise and brain neurotransmission. *Sports Med* 1995;20:160–88.
- Naidu PS, Singh A, Kaur P, Sandhir R, Kulkarni SK. Possible mechanism of action in melatonin attenuation of haloperidol-induced orofacial dyskinesia. *Pharmacol Biochem Behav* 2003;74:641–8.

- Naidu PS, Singh A, Kulkarni SK. Reversal of reserpine-induced orofacial dyskinesia and cognitive dysfunction by quercetin. *Pharmacology* 2004;70:59–67.
- Neisewander JL, Lucki I, McGonigle P. Neurochemical changes associated with the persistence of spontaneous oral dyskinesia in rats following chronic reserpine treatment. *Brain Research* 1991;558:27–35.
- Neisewander JL, Castañeda E, Davis DA. Dose-dependent differences in the development of reserpine-induced oral dyskinesia in rats: support for a model of tardive dyskinesia. *Psychopharmacology* 1994;116:79–84.
- Offen D, Gorodin S, Melamed E, Hanania J, Malik Z. Dopamine melanin is actively phagocytized by PC12 cells and cerebellar granular cells: possible implications for the etiology of Parkinson's disease. *Neurosci Lett* 1999;260:101–4.
- Ogonovszky H, Berkes I, Kumagai S, Kaneko T, Tahara S, Goto S, Radak Z. The effects of moderate, strenuous and over-training on oxidative stress markers, DNA repair, and memory, in rat brain. *Neurochem Int* 2005;46:635–40.
- Ohkawa H, Ohishi H, Yagi K. Assay for lipid peroxide in animal tissue by thiobarbituric acid reaction. *Anal Biochem* 1979;95:351–7.
- Pappolla MA, Chyan YJ, Omar RA, Hsiao K, Perry G, Smith MA, Bozner P. Evidence of oxidative stress and in vivo neurotoxicity of beta-amyloid in a transgenic mouse model of Alzheimer's disease: a chronic oxidative paradigm for testing antioxidant therapies in vivo. *Am J Pathol* 1998;152:871–7.
- Paris I, Dagnino-Subiabre A, Marcelain K, Bennett LB, Caviedes P, Caviedes R, et al. Copper neurotoxicity is dependent on dopamine-mediated copper uptake and one-electron reduction of aminochrome in a rat substantia nigra neuronal cell line. *J Neurochem* 2001;77:519–29.
- Paris I, Martinez-Alvarado P, Cardenas S, Perez-Pastene C, Graumann R, Fuentes P, et al. Dopamine-dependent iron toxicity in cells derived from rat hypothalamus. *Chem Res Toxicol* 2005;18:415–9.
- Peixoto MF, Araujo NP, Silva RH, Castro JPMV, Fukushiro DF, Faria RR, et al. Effects of gabaergic drugs on reserpine-induced oral dyskinesia. *Behav Brain Res* 2005;160:51–9.
- Petibois C, Cazorla G, Poortmans JR, Deleris G. Biochemical aspects of overtraining in endurance sports: the metabolism alteration process syndrome. *Sports Med* 2003;33:83–94.
- Radak Z, Kaneko T, Tahara S, Nakamoto H, Pucsok J, Sasvari M, et al. Regular exercise improves cognitive function and decreases oxidative damage in rat brain. *Neurochem Int* 2001;38:17–23.
- Radak Z, Toldy A, Szabo Z, Siamilis S, Nyakas C, Silye G, et al. The effects of training and detraining on memory, neurotrophins and oxidative stress markers in rat brain. *Neurochem Int* 2006;49:387–92.
- Raghavendra V, Naidu PS, Kulkarni SK. Reversal of reserpine-induced vacuous chewing movements in rats by melatonin: involvement of peripheral benzodiazepine receptors. *Brain Res* 2001;904:149–52.
- Reid MB, Haak KE, Franchek KM, Valberg PA, Kobzik L, West MS. Reactive oxygen in skeletal muscle. I. Intracellular oxidant kinetics and fatigue in vitro. *J Appl Physiol* 1992;73:1797–804.
- Sagara Y. Induction of reactive oxygen species in neurons by haloperidol. *J Neurochem* 1998;71:1002–12.
- Sarbadhikari SN. A neural network confirms that physical exercise reverses EEG changes in depressed rats. *Med Eng Phys* 1995;17:579–82.
- Sasco AJ, Paffenbarger Jr RS, Gendre I, Wing AL. The role of physical exercise in the occurrence of Parkinson's disease. *Arch Neurol* 1992;49:360–5.
- Sastre J, Asensi M, Gasco E, Pallardo FV, Ferrero JA, Furukawa T, Vina J. Exhaustive physical exercise causes oxidation of glutathione status in blood: prevention by antioxidant administration. *Am J Physiol* 1992;263:992–5.
- Segura-Aguilar J, Diaz-Veliz G, Mora S, Herrera-Marschitz M. Inhibition of DT-diaphorase is a requirement for Mn³⁺ to produce a 6-OH-dopamine like rotational behaviour. *Neurotoxicol Res* 2002;4:127–31.
- Setkovic Z, Mazur A. Physical training decreases susceptibility to subsequent pilocarpine-induced seizures in the rat. *Epil Res* 2006;71:142–8.
- Siegel S. Nonparametric statistic for the behavioral sciences. New York: McGraw-Hill; 1956. p. 117–27.
- Singh A, Naidu PS, Kulkarni SK. Possible antioxidant and neuroprotective mechanisms of FK506 in attenuating haloperidol-induced orofacial dyskinesia. *Eur J Pharmacol* 2003;477:87–94.
- Sjodin BY, Heesten WY, Apple RS. Biochemical mechanisms for oxygen free radical formation during exercise. *Sports Med* 1990;10:236–54.
- Smith AD, Zigmond MJ. Can the brain be protected through exercise? Lessons from an animal model of parkinsonism. *Exp Neurol* 2003;184:31–9.
- Somani SM, Ravi R, Rybak LP. Effect of exercise training on antioxidant system in brain regions of rat. *Pharmacol Biochem Behav* 1995;50:635–9.
- Sussman AN, Tran-Nguyen LTL, Neisewander JL. Acute reserpine administration elicits long-term spontaneous oral dyskinesia. *Eur J Pharmacol* 1997;337:157–60.
- Sutoo D, Akiyama K. Regulation of brain function by exercise. *Neurobiol Dis* 2003;13:1–14.
- Suzuki M, Katamine S, Tatsumi S. Exercise-induced enhancement of lipid peroxide metabolism in tissues and their transference into the brain in rat. *J Nutr Sci Vitaminol* 1983;29:141–51.
- Teixeira AM, Trevizol F, Colpo G, Garcia SC, Charão M, Pereira RP, et al. Influence of chronic exercise on reserpine-induced oxidative stress in rats: behavioral and antioxidant evaluations. *Pharmacol Biochem Behav* 2008;88:465–72.
- Tharp GD, Carson WH. Emotionality changes in rats following chronic exercise. *Med Sci Sports Exer* 1975;7:123–6.
- Toldy A, Stadler K, Sasvári M, Jakus J, Jung KJ, Chung HY, et al. The effect of exercise and nettle supplementation on oxidative stress markers in the rat brain. *Brain Res Bull* 2005;65:487–93.
- Tsai G, Goff DC, Chang RW, Flood J, Baer L, Coyle JT. Markers of glutamatergic neurotransmission and oxidative stress associated with tardive dyskinesia. *Am J Psychiatry* 1998;155:1207–13.
- Ziv I, Melamed E, Nardi N, Luria D, Achiron A, Offen D, Barzilai A. Dopamine induces apoptosis like cell death in cultured sympathetic neurons: a possible novel pathogenic mechanism in Parkinson's disease. *Neurosci Lett* 1994;170:136–40.